Tetrahedron 67 (2011) 210-215

Contents lists available at ScienceDirect

Tetrahedron

journal homepage: www.elsevier.com/locate/tet

Aza variant of intramolecular nucleophile-catalyzed aldol lactonization (NCAL): formal synthesis of (3*S*,4*R*) and (3*R*,4*S*) 4-(hydroxymethyl)pyrrolidin-3-ol

Dimpy Sikriwal, Dinesh K. Dikshit*

Division of Medicinal and Process Chemistry, Central Drug Research Institute (C.S.I.R.), Lucknow 226 001, UP, India

A R T I C L E I N F O

ABSTRACT

Article history: Received 2 October 2010 Received in revised form 25 October 2010 Accepted 27 October 2010 Available online 2 November 2010

Keywords: Aza sugars Aza-NCAL reaction β-Lactone O-Acetylquinidine O-Acetylquinine Aza variant of intramolecular catalytic, asymmetric nucleophile-catalyzed, aldol lactonization (NCAL) reaction has been explored to synthesize β -lactone fused nitrogen heterocycles as aza sugars' precursors by employing achiral amino acids. The utility of these bicyclic β -lactones is presented by the formal synthesis of aza sugars, (3*S*,4*R*) and (3*R*,4*S*) 4-(hydroxymethyl)pyrrolidin-3-ol.

© 2010 Elsevier Ltd. All rights reserved.

1. Introduction

Polyhydroxylated pyrrolidines, piperidines (often called amino or aza sugars), and their synthetic analogs have attracted considerable interest due to their ability to mimic sugars.¹ Their often potent inhibitory activity toward glycosidases and glycosyltransferases of various species makes them useful in a wide range of potential therapeutic areas, such as viral infections,² cancer,³ diabetes,⁴ tuberculosis,⁵ lysosomal storage diseases,⁶ and parasitic protozoa.⁷ Hydroxylated-piperidine structural framework is found in a wide variety of bio-actives, such as *cis*-3-hydroxypipecolic acid as a constituent of antibiotic tetrazomine,⁸ isofebrifugine,⁹ and 1-deoxygalactonojirimycin.^{6,10} On the other hand, *cis*-3-hydroxyproline structural motif is a part of cyclothialidine¹¹ (a potent DNA-gyrase inhibitor), slaframine,¹² castanospermine,¹³ and detoxinine.¹⁴ For the synthesis of cis-3-hydroxy-2-hydroxymethylpyrrolidines (the 'azaDNA' analog) there are only a few syntheses reported in the literature.¹⁵ In recent years, 4-hydroxymethyl-pyrrolidin-3-ol, a precursor for BCX-4208 and its analogs, has been the target of several synthetic approaches of which many are multi-steps or selectivity deficient (Fig. 1).¹⁶ It is therefore necessary to develop new short routes to this class of compounds.

Derivatives of cinchona alkaloids have shown great promise as catalyst for a broad range of asymmetric transformations thereby



Fig. 1. Structures of aza sugars and related molecules.

providing access to chiral products of high enantiopurity.¹⁷ Recently, Romo et al. have reported a high enantioselective synthesis of a series of bicyclic β -lactone fused carbocycles,¹⁸ pyrrolidinone^{19a} or tetrahydrofurans^{19b} via intramolecular nucleophile-catalyzed aldol lactonization (NCAL) of aldehyde/keto acids (Fig. 2). Inspired





 $^{^{*}}$ Corresponding author. Fax: +91 522 262 3405; e-mail address: dk_dikshit@ cdri.res.in (D.K. Dikshit).

^{0040-4020/\$ —} see front matter @ 2010 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2010.10.085



Fig. 2. Retrosynthetic analysis.

by this work, we thought it worthwhile to develop the general synthetic strategy toward the chiral synthesis of several *cis*-aza sugars and related molecules as depicted in Fig. 2.

2. Results and discussion

Achiral amino acids (3a-d) required as the starting point for NCAL reactions were prepared either by N-allylation of Cbz-AlaOH $(1a^{20} \rightarrow 2a)$ or Cbz-GABA $(1b^{20} \rightarrow 2b)$ followed by ozonolysis (route A) to afford **3a,b** or by a three-step procedure comprising (i) N- ω -alkenylation of glycine methyl ester²¹ (1c,d), (ii) one pot Cbz protection, hydrolysis of ester group (2c,d), and (iii) subsequent ozonolysis (route B), to give aldehyde-acids **3c,d** (Scheme 1). The



Scheme 1. Synthesis of aldehyde-acids 3a-d.

scope of intramolecular aza-NCAL reaction was studied using substrates 3a-d and Mukaiyama's reagent 4a under conditions as described by Romo et al.^{18a} The reactions were performed by slow syringe pump addition of aldehyde-acids **3a-d** over a period of 10 h to a magnetically stirred solution of pyridinium salt 4a and TEA in acetonitrile at room temperature (method A). The results are summarised in Table 1. It was found that the reactions with 3a and **3b** afforded racemic β -lactones, **5a** and **5b** in 53% and 51% yield, respectively (Table 1, entries 1 and 2). However, the reaction with aldehyde-acid 3c gave only the decomposed product (Table 1, entry 3).²² Use of *N*-methyl-2-bromopyridinium triflate $(4b)^{18b}$ (method B) and N-ethyl-2-bromopyridinium tetrafluoroborate (4c) (method C) greatly improved efficiency of the reaction and better yields of the cyclised products were obtained (Table 1, entries 1-4). These results were in conformity with the observations of Romo et al.^{18b}

 Table 1

 Optimization of the racemic aza-NCAL reactions







^a Substrates **3a–d** were added by syringe pump. Method A: **4a** (3.0 equiv), addition time (10 h). Method B: **4b** (3.0 equiv), addition time (3 h). Method C: **4c** (3.0 equiv), addition time (3 h).

^b Isolated yields.

^c *N*-Benzyloxycarbonyl-4,5-dihydro-1*H*-pyrrole-2-carboxylic acid was isolated albeit in low yields.

Having optimized the reaction conditions, asymmetric NCAL reactions of substrates **3a**–**c** using chiral amine catalysts were studied next (Scheme 2). It was observed that slow addition of aldehyde-acids **3a**–**c** to a solution of *O*-acetylquinidine,²³ Mukaiyama's reagent **4b**, and DIPEA in CH₃CN gave the enantiomeric bicyclic lactones, (+)-**5a** (65%), (+)-**5b** (63%), and (–)-**5c** (51%) with an ee of 92, 97, and 91%, respectively, as determined by chiral HPLC. Likewise the enantiomeric bicyclic lactones (–)-**5a** (66%), (–)-**5b** (67%), and (+)-**5c** (53%) were synthesized, using *O*-acetylquinine²⁴ with an ee of 83, 88, and 95%, respectively (Scheme 2).

The (15,55) configuration of bicyclic lactone (+)-**5a** was confirmed by its reduction with DIBAl-H to the known Cbz protected (35,4R) 4-(hydroxymethyl)pyrrolidin-3-ol [(+)-**6a**] in 43% yield,



Scheme 2. Catalytic, asymmetric aza-NCAL reactions.

which was compared with the authentic sample ($[\alpha]_{D}^{29} + 2.01$ (*c* 0.71, MeOH)) [lit.²⁵ $[\alpha]_{D}^{25} + 2.6$ (*c* 0.835, MeOH)]. Likewise, diol (–)-**6a** could also be obtained from lactone (–)-**5a** with an overall yield of 45%, which has not been reported in literature so far ($[\alpha]_{D}^{29} - 5.00$ (*c* 0.69, MeOH)) (Scheme 3).



Scheme 3. Synthesis of diols (+)-6a and (-)-6a.

3. Conclusion

In conclusion, we have delineated a formal NCAL strategy for the chiral synthesis of β -lactone fused nitrogen heterocycles where the reactive β -lactone provides a handle for further manipulations. We are currently investigating the scope of this methodology for the synthesis of natural products.

4. Experimental section

4.1. General

All NCAL reactions were done using flame-dried glassware under nitrogen atmosphere. Acetonitrile (CH₃CN), triethylamine (TEA), *N*,*N*-diisopropylethylamine (DIPEA), and dichloromethane (CH₂Cl₂) were distilled over calcium hydride. Reactions were monitored by thin layer chromatography (TLC) using 0.25 mm E. Merck pre-coated (Merch 60 F₂₅₄) silica gel plates and using ninhydrine or KMnO₄ as visualizing agent. Purification was performed by flash chromatography using silica gel (230–400 mesh). NMR spectra were recorded on a Bruker Advance-300 spectrometer. Chemical shifts are reported as parts per million (δ) relative to TMS as internal standard. Mass spectra were recorded on LCQ Advantage MAX (ESI) and JOEL JMS-600H (EI/HRMS) mass spectrometers. IR spectra were recorded on a Perkin–Elmer FT-IR RXI spectrometer. Microanalytical data were obtained using Vario-EL-III elemental analyzer. Optical rotations were determined on an Autopol III polarimeter. Chiral HPLC analyses were performed using Daicel Chiralpak IA column.

4.2. General procedure for N-allylation (2a,b)

4.2.1. N-Allyl-N-benzyloxycarbonyl- β -alanine (**2a**). To a cooled (0 °C) suspended stirred solution of NaH (4.48 g, 112.1 mmol, 60% NaH in mineral oil) in anhydrous DMF (25 ml) was added a solution of $1a^{20}$ (5.0 g, 22.4 mmol) in DMF (25 ml) dropwise under nitrogen. Once the hydrogen evolution ceased, allyl bromide (2.9 ml, 33.6 mmol) was added dropwise into the above heterogeneous solution and stirring was continued for additional 2 h at 0 °C. The reaction mixture was quenched at same temperature by the addition of 1 N HCl to become acidic (pH 2), diluted with water (250 ml), and extracted with ethyl acetate $(3 \times 80 \text{ ml})$. The combined organic extracts were washed with brine (2×40 ml), dried (Na₂SO₄), and concentrated in vacuo. The resulting crude oil was purified by flash chromatography over silica gel using ethyl acetate/hexane (3:7) to afford 2a as clear oil (4.76 g, 80.81%) [found: C, 63.58; H, 6.39; N, 5.12. C₁₃H₁₅NO₄ requires C, 63.87; H, 6.51; N, 5.32%]; R_f (1:1, EtOAc/ hexane) 0.25; v_{max} (neat) 3069, 2928, 1688, 1474, 1419, 1249, 1135, 1104, 1032, 990, 926 cm⁻¹; $\delta_{\rm H}$ (300 MHz, CDCl₃) 7.33 (5H, br s), 5.75-5.77 (1H, br), 5.13 (4H, br s), 3.93 (2H, br), 3.51-3.56 (2H, t, J 6.9 Hz), 2.59 (2H, br); δ_C (75 MHz, CDCl₃) 176.7, 156.1, 136.6, 133.5, 128.6, 128.1, 127.9, 117.0, 67.4, 50.5, 43.4, 36.8; MS (ESI) m/z: 264 $[M+H]^{+}$.

4.2.2. *N-Allyl-N-benzyloxycarbonyl-GABA* (**2b**). The acid was prepared from **1b**²⁰ (6.0 g, 25.2 mmol), NaH (5.05 g, 126.4 mmol, 60% NaH in mineral oil), and allyl bromide (3.28 ml, 37.9 mmol) at room temperature in 10 h. Purification by flash chromatography over silica gel using ethyl acetate/hexane (1:4) afforded **2b** as clear oil (4.8 g, 68.57%) [found: C, 64.73; H, 6.96; N, 5.00. C₁₅H₁₉NO₄ requires C, 64.97; H, 6.91; N, 5.05%]; *R*_f (3:2, EtOAc/hexane) 0.3; ν_{max} (neat) 3020, 2360, 1692, 1597, 1472, 1422, 1216, 1045 cm⁻¹; δ_{H} (300 MHz, CDCl₃) 7.32–7.34 (5H, br m), 5.76 (1H, br), 5.13 (4H, br s), 3.87 (2H, br), 3.31 (2H, br), 2.34 (2H, br), 1.86 (2H, br); δ_{C} (75 MHz, CDCl₃) 178.2, 156.4, 136.7, 133.6, 128.5, 128.1, 127.9, 117.0, 67.4, 49.7, 46.3, 31.2, 23.3; MS (ESI) *m/z*: 278 [M+H]⁺.

4.3. General procedure for mono N-alkenylation (1c,d)

4.3.1. Methyl N-(but-3-enyl)glycinate (**1c**). K₂CO₃ (9.39 g, 67.9 mmol) was added to a solution of methyl glycinate HCl^{21} (3.15 g, 35.4 mmol) in CH₃CN (150 ml) and the mixture was stirred for 1 h. 4-Bromo-1-butene (3.0 ml, 29.5 mmol) was added to the mixture and the reaction mixture was stirred for 48 h. The insoluble materials

were filtered off and the filtrate was concentrated under reduced pressure. To the residue was added water (150 ml) and extracted with ethyl acetate (3×60 ml). The combined organic extracts were washed with brine (2×25 ml), dried (Na₂SO₄), and concentrated in vacuo to afford a clear liquid. Purification by flash chromatography on silica gel using ethyl acetate/hexane (4:6 \rightarrow 10:0) furnished **1c** as clear oil (3.1 g, 73.28%) [found: C, 58.82; H, 9.10; N, 9.81. C₇H₁₃NO₂ requires C, 58.72; H, 9.15; N, 9.78%]; *R*_f (EtOAc) 0.3; *v*_{max} (neat) 3020, 1743, 1634, 1437, 1215 cm⁻¹; $\delta_{\rm H}$ (300 MHz, CDCl₃) 5.74–5.88 (1H, m), 5.05–5.15 (2H, m), 3.74 (3H, s), 3.44 (3H, s), 2.67–2.72 (2H, t, *J* 6.9 Hz,), 2.24–2.31 (2H, q, *J* 6.9, 13.8 Hz); $\delta_{\rm C}$ (75 MHz, CDCl₃) 172.9, 136.1, 166.6, 51.8, 50.7, 48.5, 34.3; MS (ESI) *m/z*: 144 [M+H]⁺.

4.3.2. *Methyl N*-(*pent-3-enyl*)*glycinate* (**1d**). Compound **1d** was prepared from methyl glycinate ·HCl (3.15 g, 35.4 mmol), K₂CO₃ (9.39 g, 67.9 mmol), and 5-bromo-1-pentene (3.5 ml, 29.5 mmol). Purification by flash column chromatography on silica gel using chloroform as eluant isolated **1d** as clear oil (3.16 g, 68.10%) [found: C, 61.01; H, 9.28; N, 8.93. C₈H₁₅NO₂ requires C, 61.12; H, 9.62; N, 8.91%]; *R*_f (CHCl₃) 0.3; *v*_{max} (neat) 3020, 2933, 1740, 1639, 1439, 1217 cm⁻¹; $\delta_{\rm H}$ (300 MHz, CDCl₃) 5.76–5.89 (1H, m), 4.96–5.07 (2H, m), 3.74 (3H, s), 3.42 (3H, s), 2.60–2.65 (2H, t, *J* 7.1 Hz), 2.08–2.15 (2H, q, *J* 7.1, 14.4 Hz), 1.56–1.66 (2H, m); $\delta_{\rm C}$ (75 MHz, CDCl₃) 173.1, 138.4, 114.8, 51.8, 50.8, 49.1, 31.4, 29.2; MS (ESI) *m/z*: 158 [M+H]⁺.

4.4. General procedure for Cbz introduction and hydrolysis (2c,d)

4.4.1. N-(Benzyloxycarbonyl)-N-(but-3-enyl)-glycine (2c). To a cooled (0 °C) stirred solution of 1c (2.63 g, 18.3 mmol) and TEA (5.63 ml, 40.4 mmol) in anhydrous CH₂Cl₂ (45 ml) was added Cbz-Cl (2.97 ml, 21.2 mmol) dropwise under nitrogen. The clear yellow solution was warmed to room temperature and stirred for 2 h, at which point the volatiles were removed under reduced pressure to afford yellow oil. The crude oil was then dissolved in THF/H₂O (4:1, 50 ml) with stirring followed by the addition of LiOH \cdot H₂O (2.31 g, 55.1 mmol) at room temperature. After stirring for 2 h, the reaction mixture was diluted with water (30 ml) and extracted with ether $(2 \times 35 \text{ ml})$. The pH of aqueous layer was adjusted to 2–3 by the addition of dilute hydrochloric acid (1:1) at 0 °C and extracted the liberated oil with ethyl acetate (4×50 ml). The combined organic extracts were washed with brine $(2 \times 45 \text{ ml})$ and dried (Na_2SO_4) . Removal of the solvent in vacuo gave the analytically pure product 2c as clear oil (4.38 g, 90.68%) [found: C, 63.76; H, 6.40; N, 5.12. C14H17NO4 requires C, 63.87; H, 6.51; N, 5.32%]; Rf (7:3, EtOAc/ hexane) 0.35; *v*_{max} (neat) 3020, 2361, 1696, 1475, 1431, 1367, 1217, 1149 cm⁻¹; $\delta_{\rm H}$ (300 MHz, CDCl₃) 7.27–7.37 (5H, br m), 5.72–5.78 (1H, m), 5.15–5.19 (2H, d, / 11.4 Hz), 5.03–5.07 (2H, app t, / 6.9 Hz), 4.04-4.07 (2H, d, / 10.5 Hz), 3.43 (2H, br), 2.32 (2H, br); $\delta_{\rm C}$ (75 MHz, CDCl₃) 174.5/174.3 (rotamers), 156.8/156.0 (rotamers), 136.4/136.3 (rotamers), 135.0/134.8 (rotamers), 128.57/128.51 (rotamers), 128.1/ 128.0 (rotamers), 127.8/127.7 (rotamers), 117.2/117.0 (rotamers), 67.7/67.5 (rotamers), 49.4/48.9 (rotamers), 48.4/48.0 (rotamers), 32.9/32.4 (rotamers); MS (ESI) *m*/*z*: 264 [M+H]⁺.

4.4.2. *N*-(*Benzyloxycarbonyl*)-*N*-(*pent*-3-*enyl*)-*glycine* (**2d**). Clear oil (78.49%) [found: C, 64.73; H, 6.77; N, 4.99. $C_{15}H_{19}NO_4$ requires C, 64.97; H, 6.91; N, 5.05%]; *R*_f(1:9, MeOH/CHCl₃) 0.3; ν_{max} (neat) 2935, 2363, 1699, 1470, 1433, 1221 cm⁻¹; δ_{H} (300 MHz, CDCl₃) 7.28–7.37 (5H, m), 5.71–5.83 (1H, m), 5.15–5.19 (2H, d, *J* 11.9 Hz), 4.96–5.07 (2H, m), 4.02–4.07 (2H, d, *J* 12.6 Hz), 3.33–3.40 (2H, q, *J* 7.1 Hz), 2.01–2.12 (2H, m), 1.61–1.68 (2H, m); δ_{C} (75 MHz, CDCl₃) 175.2/174.8 (rotamers), 156.9/156.0 (rotamers), 137.8/137.6 (rotamers), 136.4, 128.6, 128.2/128.1 (rotamers), 127.9/127.8 (rotamers), 148.5/48.0

(rotamers), 30.9/30.8 (rotamers), 27.5/27.1 (rotamers); MS (ESI) *m/z*: 278 [M+H]⁺.

4.5. General procedure for ozonolysis (3a–d)

Ozone was bubbled through a cooled (-78 °C) solution of **2a** (3.2 g, 11.0 mmol) in 75 ml of anhydrous CH₂Cl₂ for 1 h. After that, a stream of argon was passed through the cooled solution for 30 min to eliminate the excess of ozone. The cooled reaction mixture was then quenched with excess of Me₂S (3.57 ml, 48.6 mmol), allowed to warm up to room temperature and treated with cold water (40 ml), extracted with CH₂Cl₂ (3×45 ml), dried (Na₂SO₄), and concentrated in vacuo to obtain the aldehyde **3a** (2.59 g, 80.4%). Similarly **3b–d** were obtained from **2b–d** in 81–82% and were used immediately for NCAL reaction without any further purification.

4.6. *N*-Methyl-2-bromopyridinium triflate (4b)^{18b}

Pyridinium salt **4b** was prepared from 2-bromopyridine (2.0 ml, 20.9 mmol) and methyl trifluoromethanesulfonate (2.3 ml, 20.9 mmol) in CH₂Cl₂ (10 ml) according to the literature method as white crystalline solid (6.46 g, 95.98%); mp 158–160 °C; ν_{max} (KBr) 3087, 1616, 1491, 1443, 1267, 1156, 1032 cm⁻¹; $\delta_{\rm H}$ (300 MHz, CDCl₃+CD₃OD) 9.10–9.12 (1H, d, *J* 5.2 Hz), 8.34–8.40 (1H, m), 8.27–8.30 (1H, d, *J* 7.2 Hz), 8.0–8.05 (1H, m), 4.48 (3H, s); $\delta_{\rm C}$ (75 MHz, CDCl₃) 149.0, 146.3, 133.9, 127.1, 122.5, 118.2, 50.8.

4.7. General procedure for the racemic aza-NCAL reaction as described for (\pm) -5a

4.7.1. Benzyl 7-oxo-6-oxa-3-azabicyclo[3.2.0]heptane-3-carboxylate $[(\pm)-5a]$. Method B: To a stirred solution of Mukaiyama's reagent 4b (1.65 g, 5.13 mmol, 3.0 equiv) and TEA (0.95 ml, 6.85 mmol, 4.0 equiv) in CH₃CN (30 ml) at 25 °C was added via syringe pump a solution of aldehyde-acid **3a** (0.45 g, 1.71 mmol) in CH₃CN (20 ml) over a period of 3 h. Stirring of the resulting dark red solution was continued for another 12 h. The volatiles were removed under reduced pressure and to the crude reaction mixture were added ethyl acetate (150 ml) and saturated aqueous NH₄Cl (150 ml). The phases were separated, and the aqueous layer was extracted with ethyl acetate $(2 \times 50 \text{ ml})$. The combined organic phases were washed with brine (100 ml), dried (Na₂SO₄), filtered, and concentrated to afford a brown residue. Purification of the crude residue by flash chromatography on SiO₂ using ethyl acetate/hexane (13:7) afforded the β -lactone-**5a** as light yellow oil (0.7 g, 76.07%) [found: C, 63.23; H, 5.41; N, 5.65. C₁₃H₁₃NO₄ requires C, 63.15; H, 5.30; N, 5.67%]; R_f (7:3, EtOAc/hexane) 0.35; $v_{\rm max}$ (neat) 1836, 1705, 1423, 1356, 1261, 1227, 1187, 1114 cm $^{-1}$; $\delta_{\rm H}$ (300 MHz, CDCl₃) 7.32–7.40 (5H, m, ArH), 5.15 (2H, br s, PhCH₂), 5.09-5.12 (1H, dd, / 4.0, 5.9 Hz, H-5), 4.23-4.28 (2H, br m, H-2+H-4), 4.08-4.12 (1H, m, H-1), 3.29-3.35 (2H, m, H-2'+H-4'); $\delta_{\rm C}$ (75 MHz, CDCl₃) 168.6 (C=0, lactone), 155.0 (C=0, carbamate), 136.1 (qC), 128.5, 128.2, 128.0 (ArC), 74.9/74.3 (C-5, rotamers), 67.5 (PhCH₂), 56.1/55.4 (C-1, rotamers), 49.6 (C-4), 45.6 (C-2); HRMS (ESI): [M+H]⁺ found 248.0919. C₁₃H₁₄NO₄ requires 248.0922.

4.7.2. Benzyl 7-oxo-8-oxa-3-azabicyclo[4.2.0]octane-3-carboxylate $[(\pm)$ -**5b**]. This lactone was prepared from oxo-acid **3b** (0.286 g, 1.02 mmol), pyridinium salt **4b** (0.989 g, 3.07 mmol), and TEA (0.49 ml, 3.55 mmol). Purified by flash column chromatography using ethyl acetate/CH₂Cl₂ (7:93) as eluant isolated **5b** as clear viscous oil (0.19 g, 71.16%) [found: C, 64.24; H, 5.81; N, 5.34. C₁₄H₁₅NO₄ requires C, 64.36; H, 5.79; N, 5.36%]; *R*_f (1:9, EtOAc/CH₂Cl₂) 0.5; *v*_{max} (neat) 1826, 1698, 1421, 1353, 1290, 1224, 1117,

1052 cm⁻¹; $\delta_{\rm H}$ (300 MHz, CDCl₃) 7.34 (5H, br s, Ar*H*), 5.08–5.21 (2H, m, PhCH₂), 4.74–4.80 (1H, br d, *J* 15.6 Hz, *H*-1), 4.42–4.47 (0.5H, d, *J* 15.6 Hz, *H*-2, rotamers), 4.32–4.37 (0.5H, d, *J* 15.6 Hz, *H*-2, rotamers), 3.82–3.87 (1H, m, *H*-6), 3.67–3.69 (1H, br, *H*-4), 3.46–3.56 (1H, td, *J* 4.5, 12.6 Hz, *H*-4'), 3.36–3.41 (1H, d, *J* 15.6 Hz, *H*-2'), 2.09–2.18 (1H, app t, *J* 15.5 Hz, *H*-5), 1.89–2.02 (1H, m, *H*-5'); $\delta_{\rm C}$ (75 MHz, CDCl₃) 169.6 (C=O, lactone), 155.8 (C=O, carbamate), 136.4 (qC), 128.6, 128.2, 127.9 (ArC), 69.1/68.7 (C-1, rotamers), 67.5 (PhCH₂), 47.6 (C-6), 42.0/41.5 (C-2, rotamers), 39.9 (C-4), 19.7 (C-5); HRMS (ESI): [M+H]⁺ found 262.1080. C₁₄H₁₆NO₄ requires 262.1079.

4.7.3. Benzyl 7-oxo-8-oxa-2-azabicyclo[4.2.0]octane-2-carboxylate $[(\pm)-5c]$. This lactone was prepared from oxo-acid 3c (0.225 g, 0.84 mmol), pyridinium salt 4b (0.819 g, 2.54 mmol), and TEA (0.47 ml, 3.39 mmol). Purified by flash column chromatography using ethyl acetate/CH₂Cl₂ (1:49) as eluant isolated **5c** as viscous oil (0.115 g, 55.02%) [found: C, 63.05; H, 5.39; N, 5.61. C₁₃H₁₃NO₄ requires C, 63.15; H, 5.30; N, 5.67%]; R_f (1:9, EtOAc/CH₂Cl₂) 0.6; v_{max} (neat) 1837, 1708, 1422, 1347, 1306, 1266, 1216, 1108, 1059 cm⁻¹; $\delta_{\rm H}$ (300 MHz, CDCl₃) 7.32-7.36 (5H, m, ArH), 5.53-5.65 (1H, br, H-1), 5.15–5.17 (3H, m, PhCH₂+H-5), 4.12–4.19 (1H, app t, J 9.7 Hz, H-3), 3.37-3.46 (1H, td, / 6.2, 11.5 Hz, H-3'), 2.29-2.36 (1H, dd, / 6.2, 14.8 Hz, H-4), 1.87–2.00 (1H, m, H-4'); δ_C (75 MHz, CDCl₃) 166.7 (C=O, lactone), 153.3 (C=O, carbamate), 135.9 (qC), 128.7, 128.4, 128.2 (ArC), 77.8 (C-5), 70.2/69.8 (C-1, rotamers), 67.9 (PhCH₂), 44.0 (C-3), 29.1 (C-4); HRMS (ESI): [M+Na]⁺ found 270.0741. C₁₃H₁₃NNaO₄ requires 270.0737.

4.7.4. Benzyl 8-oxo-7-oxa-2-azabicyclo[4.2.0]octane-2-carboxylate $[(\pm)-5d]$. This lactone was prepared from oxo-acid 3d (0.41 g, 1.46 mmol), pyridinium salt 4b (1.418 g, 4.4 mmol), and TEA (0.81 ml, 5.87 mmol). Purified by flash column chromatography using ethyl acetate/hexane (3:7) as eluant isolated 5d as viscous oil (0.202 g, 52.74%) [found: C, 64.29; H, 5.65; N, 5.39. C₁₄H₁₅NO₄ requires C, 64.36; H, 5.79; N, 5.36]; R_f (1:1, EtOAc/hexane) 0.45; $v_{\rm max}$ (neat) 1832, 1703, 1418, 1310, 1216, 1114, 1037 cm⁻¹; $\delta_{\rm H}$ (300 MHz, CDCl₃) 7.35 (5H, br s, ArH), 5.80–5.82 (0.5H, d, J 6.6 Hz, H-1, rotamers), 5.58-5.60 (0.5H, d, J 6.6 Hz, H-1, rotamers), 5.12-5.22 (2H, m, PhCH₂), 4.97 (1H, br s, H-6), 3.68-3.72 (1H, dd, J 3.6, 11.8 Hz, H-3), 3.36-3.41 (1H, m, H-3'), 2.26-2.30 (1H, d, J 12.6 Hz, H-4), 1.80–1.99 (3H, m, H-5+H-5'+H-4'); δ_C (75 MHz, CDCl₃) 170.5/169.9 (C=O, lactone, rotamers), 155.7/154.6 (C=O, carbamate, rotamers), 136.0 (qC), 128.6, 128.4, 128.1 (ArC), 72.1/71.9 (C-6, rotamers), 68.1/68.0 (PhCH₂, rotamers), 59.7/59.5 (C-1, rotamers), 43.0/42.9 (C-3, rotamers), 25.9 (C-4), 16.1/15.8 (C-5, rotamers); HRMS (ESI): [M+H]⁺ found 262.1094. C₁₄H₁₆NO₄ requires 262.1079.

4.8. General procedure for asymmetric aza-NCAL reaction as described for β -lactone-(+)-5a

4.8.1. (15,55)-Benzyl 7-oxo-6-oxa-3-azabicyclo[3.2.0]heptane-3-carboxylate [(+)-**5a**]. To a stirred solution of O-acetylquinidine²³ (71 mg, 0.196 mmol), Mukaiyama's reagent **4b** (1.895 g, 5.88 mmol), and DIPEA (1.36 ml, 7.84 mmol) in CH₃CN (35 ml) was added a solution of aldehyde-acid **3a** (0.52 g, 1.96 mmol) in CH₃CN (30 ml) via syringe pump over a period of 3 h. After the addition was completed, the reaction was stirred for additional 30 h at 25 °C. The solvent was removed in vacuo, and the residue was partitioned between ethyl acetate (150 ml) and saturated NH₄Cl (100 ml). The phases were separated, and the aqueous layer was extracted with ethyl acetate (2×50 ml). The combined organic phases were washed with brine (100 ml), dried (Na₂SO₄), and concentrated in vacuo. Purification of the crude residue by flash chromatography on SiO₂ using ethyl acetate/hexane (13:7) afforded the β -lactone-(+)-**5a** as light yellow oil (0.315 g, 64.94%). [α] $_{D}^{29}$ +111.4 (*c* 0.20, CHCl₃). Enantiomeric excess (ee) was determined to be 92% by chiral stationary phase HPLC analysis using Daicel Chiralpak IA column (MtBE/EtOH=99:1, flow rate 0.6 ml/min, λ_{max} 213.9 nm), $t_{15,55}$ =20.74 min (major), $t_{1R,5R}$ =22.66 min (minor). All other spectroscopic data matched that displayed by (±)-**5a**.

4.8.2. (15,6S)-Benzyl 7-oxo-8-oxa-3-azabicyclo[4.2.0]octane-3-carboxylate [(+)-**5b**]. This lactone was prepared from oxo-acid **3b** (0.25 g, 0.89 mmol), pyridinium salt **4b** (0.864 g, 2.68 mmol), DIPEA (0.62 ml, 3.58 mmol), and O-acetylquinidine (32 mg, 0.089 mmol). Purification by flash column chromatography on silica gel using ethyl acetate/CH₂Cl₂ (7:93) as eluant gave (+)-**5b** as viscous oil (0.147 g, 63.09%). [α]_D²⁹+118.7 (*c* 1.09, MeOH). Enantiomeric excess (ee) was determined to be 97% by chiral stationary phase HPLC analysis using Daicel Chiralpak IA column (MtBE/EtOH=99:1, flow rate 0.6 ml/min, λ_{max} 209 nm), $t_{15,65}$ =20.98 min (major), $t_{1R,6R}$ =32.31 min (minor). All other spectroscopic data matched that displayed by (±)-**5b**.

4.8.3. (15,5R)-Benzyl 7-oxo-8-oxa-2-azabicyclo[4.2.0]octane-2-carboxylate [(-)-**5c**]. This lactone was prepared from oxo-acid **3c** (0.29 g, 1.09 mmol), pyridinium salt **4b** (1.05 g, 3.27 mmol), DIPEA (0.76 ml, 4.37 mmol), and O-acetylquinidine (40 mg, 0.109 mmol). Purification by flash column chromatography on silica using ethyl acetate/CH₂Cl₂ (1:49) as eluant gave (-)-**5c** as viscous oil (0.137 g, 50.74%). [α]_D²⁷ -132.4 (*c* 0.16, MeOH). Enantiomeric excess (ee) was determined to be 91% by chiral stationary phase HPLC analysis using Daicel Chiralpak IA column (MtBE/EtOH=98:2, flow rate 0.8 ml/min, λ_{max} 213 nm), $t_{1R,55}$ =8.82 min (minor), $t_{15,5R}$ =11.64 min (major). All other spectroscopic data matched that of the racemic compound (±)-**5c**.

4.8.4. (1*R*,5*R*)-Benzyl 7-oxo-6-oxa-3-azabicyclo[3.2.0]heptane-3-carboxylate [(-)-**5a**]. The lactone was prepared from aldehyde-acid **3a** (0.348 g, 1.31 mmol), pyridinium salt **4b** (1.268 g, 3.93 mmol), DIPEA (0.91 ml, 5.25 mmol), and O-acetylquinine²⁴ (48 mg, 0.13 mmol) following the procedure as described for (+)-**5a**. Purification by flash chromatography on SiO₂ using ethyl acetate/ hexane (13:7) gave the (-)-**5a** as colorless oil (0.214 g, 66.04% yield). [α]_D⁵⁹ -39.1° (*c* 0.44, CHCl₃). Enantiomeric excess (ee) was determined to be 83% by chiral stationary phase HPLC analysis using Daicel Chiralpak IA column (MtBE/EtOH=99:1, flow rate 0.6 ml/min, λ_{max} 213.9 nm), $t_{15,55}$ =20.56 min (minor), $t_{1R,5R}$ = 22.42 min (major). All other spectroscopic data matched with (±)-**5a**.

4.8.5. (1R,6R)-Benzyl 7-oxo-8-oxa-3-azabicyclo[4.2.0]octane-3-carboxylate [(-)-**5b**]. This lactone was prepared from oxo-acid **3b** (0.195 g, 0.698 mmol), pyridinium salt **4b** (0.674 g, 2.09 mmol), DIPEA (0.48 ml, 2.79 mmol), and O-acetylquinine (25 mg, 0.069 mmol). Purification by flash column chromatography on silica gel using ethyl acetate/CH₂Cl₂ (7:93) as eluant gave (-)-**5b** as viscous oil (0.12 g, 67.03%). [α]_D²⁹ –63.3 (*c* 1.11, MeOH). Enantiomeric excess (ee) was determined to be 88% by chiral stationary phase HPLC analysis using Daicel Chiralpak IA column (MtBE/EtOH=99:1, flow rate 0.6 ml/min, λ_{max} 209 nm), $t_{1S,6S}$ =20.05 min (minor), $t_{1R,6R}$ =33.36 min (major). All other spectroscopic data matched with the racemic compound (±)-**5b**.

4.8.6. (1R,5S)-Benzyl 7-oxo-8-oxa-2-azabicyclo[4.2.0]octane-2-carboxylate [(+)-**5c**]. This lactone was prepared from oxo-acid **3c** (0.2 g, 0.753 mmol), pyridinium salt **4b** (0.728 g, 2.26 mmol), DIPEA (0.52 ml, 3.01 mmol), and O-acetylquinine (27 mg, 0.075 mmol). Purification by flash column chromatography on silica using ethyl acetate/CH₂Cl₂ (1:49) as eluant gave (+)-**5c** as viscous oil (0.098 g, 52.68%). [α]_D²⁸ +67.7 (*c* 0.37, MeOH). Enantiomeric excess (ee) was determined to be 95% by chiral stationary phase HPLC analysis using Daicel Chiralpak IA column (MtBE/EtOH=98:2, flow rate 0.8 ml/min, λ_{max} 213 nm), $t_{1R,5S}$ =8.76 min (major), $t_{15,5R}$ =11.63 min (minor). All other spectroscopic data matched with the racemic compound (±)-**5c**.

4.9. General procedure for reduction as described for diol (+)-6a

4.9.1. (3S,4R)-N-Benzyloxycarbonyl-4-(hydroxymethyl)pyrrolidin-3-ol [(+)-6a]. To a magnetically stirred solution of (+)-5a (0.17 g, 0.687 mmol) in CH₂Cl₂ (5.3 ml) was added dropwise a solution of DIBAl-H (1.0 M in toluene, 0.738 ml, 0.742 mmol) at 0 °C. After 5 min, the solution was warmed to room temperature and stirred overnight. The reaction mixture was then cooled to 0 °C. diluted with ethyl acetate (2.4 ml), and quenched with acetone (1.5 ml) and Rochelle's salt (4.0 ml). The mixture was vigorously stirred at 25 °C for 10 h. The layers were separated and the aqueous layer was extracted with ethyl acetate $(2 \times 4 \text{ ml})$. The combined organic layers were washed with brine (8 ml), dried over Na₂SO₄, filtered, and concentrated in vacuo. Purification by flash chromatography on SiO₂ using MeOH/ethyl acetate (1:9) gave the diol (+)-**6a** as a colorless oil (74 mg, 43.19% yield) [found: C, 62.23; H, 6.75; N, 5.52. C₁₃H₁₇NO₄ requires C, 62.14; H, 6.82; N, 5.57%]; R_f (1:9, MeOH/ EtOAc) 0.3; $[\alpha]_D^{29}$ +2.01 (*c* 0.71, MeOH) [lit.²⁵ $[\alpha]_D^{25}$ +2.6 (*c* 0.835, MeOH)]; v_{max} (neat) 3389, 2920, 1682, 1433, 1358, 1216, 1136, 1097 cm⁻¹; δ_H (300 MHz, CDCl₃) 7.28–7.32 (5H, m, Ar*H*), 5.04–5.14 (2H, m, PhCH₂), 4.41 (1H, br s, H-3), 3.78-3.83 (2H, t, J 7.8 Hz, CH₂OH), 3.44-3.56 (3H, m, H-2+H-2'+H-5), 3.32-3.39 (1H, t, J 10.5, H-5'), 2.28-2.29 (1H, br, H-4); δ_C (75 MHz, CDCl₃) 155.5 (C=O), 136.8 (qC), 128.6, 128.1, 127.9 (ArC), 72.3/71.5 (C-3, rotamers), 67.1 (PhCH₂), 60.3 (CH₂OH), 55.2/54.9 (C-2, rotamers), 45.9/45.6 (C-5, rotamers), 45.1/44.3 (C-4, rotamers); MS (ESI) m/z: 252 [M+H]⁺; HRMS (ESI): [M+H]⁺ found 252.1216. C₁₃H₁₈NO₄ requires 252.1235.

4.9.2. (3*R*,4*S*)-*N*-Benzyloxycarbonyl-4-(hydroxymethyl)pyrrolidin-3-ol [(-)-**6a**]. The diol was prepared from lactone (-)-**5a** (0.142 g, 0.574 mmol) and DIBAl-H (1.0 M in toluene, 0.618 ml, 0.62 mmol). Purification by flash chromatography on SiO₂ using MeOH/ethyl acetate (1:9) gave the diol (-)-**6a** as colorless oil (58 mg, 44.16% yield). [α]_D²⁹ -5.0 (*c* 0.69, MeOH). All other spectroscopic data matched with diol (+)-**6a**.

Acknowledgements

D.S. thanks CSIR-UGC, New Delhi, India, for the financial support in the form of Senior Research Fellowships. We thank SAIF, CDRI, for the spectral data.

Supplementary data

¹H, ¹³C, and HSQC spectral data for lactones (\pm) -**5a**–**d** and diol (+)-**6a**, chiral HPLC data for **5a**–**c**, and ¹H and ¹³C spectral data for **4b** are provided. Supplementary data associated with this article can be found in the online version, at doi:10.1016/j.tet.2010.10.085. These data include MOL files and InChIKeys of the most important compounds described in this article.

References and notes

- (a) Winchester, B.; Fleet, G. W. J. Glycobiology **1992**, *2*, 199; (b) O'Hagan, D. Nat. Prod. Rep. **1997**, *14*, 637; (c) Stütz, A. E. Iminosugars as Glycosidase Inhibitors; Wiley: Weinheim, 1999; (d) Asano, N.; Nash, R. J.; Molyneux, R. J.; Fleet, G. W. J. Tetrahedron: Asymmetry **2000**, *11*, 1645.
- (a) Karpas, A.; Fleet, G. W. J.; Dwek, R. A.; Petursson, S.; Namgoong, S. K.; Ramsden, N. G.; Jacob, G. S.; Rademacher, T. W. Proc. Natl. Acad. Sci. U.S.A. 1988, 85, 9229; (b) Block, T. M.; Xu, X. L.; Platt, F. M.; Foster, G. R.; Gerlich, W. H.; Blumberg, B. S.; Dwek, R. A. Proc. Natl. Acad. Sci. U.S.A. 1994, 91, 2235.
- 3. Goss, P. E.; Baker, M. A.; Carver, J. P.; Dennis, J. W. Clin. Cancer Res. 1995, 1, 935.
- Watson, K. A.; Mitchell, E. P.; Johnson, L. N.; Son, J. C.; Bichard, C. J. F.; Orchard, M. G.; Fleet, G. W. J.; Oikonomakos, N. G.; Leonidas, D. D.; Kontou, M.; Papageorgiou, A. C. Biochemistry 1994, 33, 5745.
- (a) Davis, B. G.; Brandstetter, T. W.; Hackett, L.; Winchester, B. G.; Nash, R. J.; Watson, A. A.; Griffiths, R. C.; Smith, C.; Fleet, G. W. J. *Tetrahedron* 1999, 55, 4489; (b) Shilvock, J. P.; Wheatley, J. R.; Nash, R. J.; Watson, A. A.; Griffiths, R. C.; Butters, T. D.; Muller, M.; Watkin, D. J.; Winkler, D. A.; Fleet, G. W. J. *J. Chem. Soc., Perkin Trans.* 1 1999, 2735.
- 6. Fan, J. Q.; Ishii, S.; Asano, N.; Suzuki, Y. Nat. Med. 1999, 5, 112.
- (a) Li, C. M.; Tyler, P. C.; Furneaux, R. H.; Kicska, G.; Xu, Y. M.; Grubmeyer, C.; Girvin, M. E.; Schramm, V. L. *Nat. Struct. Biol.* **1999**, 6, 582; (b) Miles, R. W.; Tyler, P. C.; Evans, G. B.; Furneaux, R. H.; Parkin, D. W.; Schramm, V. L. *Biochemistry* **1999**, 38, 13147.
- B. Scott, J. D.; Tippie, T. N.; Williams, R. M. Tetrahedron Lett. 1998, 39, 3659.
- (a) Kuehl, F. A., Jr.; Spencer, C. F.; Folkers, K. J. Am. Chem. Soc. 1948, 70, 2091;
 (b) Kobayashi, S.; Ueno, M.; Suzuki, R. Tetrahedron Lett. 1999, 40, 2175.
- Kato, A.; Kato, N.; Kano, E.; Adachi, I.; Ikeda, K.; Yu, L.; Okamoto, T.; Banba, Y.; Ouchi, H.; Takahata, H.; Asano, N. *J. Med. Chem.* **2005**, *48*, 2036.
 Watanabe, J.; Nakada, N.; Sawairi, S.; Shimada, H.; Ohshima, S.; Kamiyama, T.;
- Watanabe, J.; Nakada, N.; Sawairi, S.; Shimada, H.; Ohshima, S.; Kamiyama, T.; Arisawa, M. J. Antibiot. 1994, 47, 32.
- 12. Gardiner, R. A.; Rinehart, K. L.; Snyder, J. J.; Broquist, H. P. J. Am. Chem. Soc. **1968**, 90, 5639.
- Hohenschutz, L. D.; Bell, E. A.; Jewess, P. J.; Leworthy, D. P.; Pryce, R. J.; Arnold, E.; Clardy, J. Phytochemistry 1981, 20, 811.
- 14. Kakinuma, K.; Otake, N.; Yonehara, H. Tetrahedron Lett. 1972, 2509.
- (a) Mascavage, L. M.; Lu, Q.; Vey, J.; Dalton, D. R.; Carroll, P. J. J. Org. Chem. 2001, 66, 3621; (b) Yoshitomi, Y.; Makino, K.; Hamada, Y. Org. Lett. 2007, 9, 2457.
- (a) Godskesen, M.; Lundt, I. *Tetrahedron Lett.* **1998**, 39, 5841; (b) Kotian, P. L.; Chand, P. *Tetrahedron Lett.* **2005**, 46, 3327; (c) Chen, L.; Jones, E. D.; Ma, D.; Baylis, D. C.; Li, B.; Coates, J. A. V.; Xie, X.; Rhodes, D. I.; Chen, R.; Deadman, J. J. W. O. 092293A1, 2009.
- 17. For a review see: Kacprzak, K.; Gawroński, J. Synthesis 2001, 961.
- (a) Cortez, G. S.; Tennyson, R. L.; Romo, D. J. Am. Chem. Soc. 2001, 123, 7945;
 (b) Oh, S. H.; Cortez, G. S.; Romo, D. J. Org. Chem. 2005, 70, 2835;
 (c) Henry-Rivad, H.; Lee, C. S.; Purohit, V. C.; Romo, D. Org. Lett. 2006, 8, 4363.
- Riyad, H.; Lee, C. S.; Purohit, V. C.; Romo, D. Org. Lett. 2006, 8, 4363.
 19. (a) Ma, G.; Nguyen, H.; Romo, D. Org. Lett. 2007, 9, 2143; (b) Morris, K. A.; Arendt, K. M.; Oh, S. H.; Romo, D. Org. Lett. 2010, 12, 3764.
- 20. Schmuck, C.; Rehm, T.; Geiger, L.; Schäfer, M. J. Org. Chem. 2007, 72, 6162.
- Almeida, J. F.; Anaya, J.; Martin, N.; Grande, M.; Moran, J. R.; Caballero, M. C. Tetrahedron: Asymmetry 1992, 3, 1431.
- 22. N-Benzyloxycarbonyl-4,5-dihydro-1H-pyrrole-2-carboxylic acid was isolated albeit in low yields.
- 23. Waddell, T. G.; Woods, L. A.; Harrison, W.; Meyer, G. M. J. Tenn. Acad. Sci. 1984, 59, 48.
- 24. Pracejus, H.; Matje, H. J. Prakt. Chem. 1964, 195.
- 25. Asahina, Y.; Takei, M.; Kimura, T.; Fukuda, Y. J. Med. Chem. 2008, 51, 3238.